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Enclosure to letter dated 15 November 2005 concerning European Patent Appln. No. PCT/EP2005/000555; - DSM IP Assets B.V. -; ref:21726W0, clean version

REVISED CLAIMS APTI Rec'd PCT/PTO 17 JUL 2006

 Process for the biochemical synthesis of 6-amino caproic acid wherein either 6-aminohex-2-enoic acid of formula [1] (6-AHEA)

 $H_2N - CH_2 - CH_2 - CH_2 - CH = CH - COOH$

[1]

or wherein 6-amino-2-hydroxy-hexanoic acid (6-AHHA), a compound capable of being transformed into 6-aminohex-2-enoic acid, is treated with an enzyme having α,β -enoate reductase activity towards molecules containing an α,β -enoate group and a primary amino group, in particular with an enzyme having α,β -enoate reductase activity towards 6-aminohex-2-enoic acid.

- 2. Process according to claim 1, characterized in that the enzyme having α;β-enoate reductase activity is an enzyme originating from a microorganism from the group of species of Acetobacterium sp., Acremonium sp., Agrobacterium sp., Burkholderia sp., Cephalosporium sp., Clostridium sp., Escherichia sp., Moorella sp., Ochrobactrum sp., Pseudomonas sp., Salmonella sp., Shigella sp., Tilachlidium sp., Yersinia sp., and Vibrio sp.
- 3. Process according to one of claims 1 or 2, characterized in that the enzyme having α,β-enoate reductase activity is an enzyme originating from *Acremonium* sp., *Clostridium* sp., *Moorella* sp. or *Ochrobactrum* sp.
- 4. Process according to claim 3, characterized in that the enzyme having is an enzyme from Acremonium strictum CBS114157, Clostridium tyrobutyricum DSM1460, Moorella thermoacetica DSM1974, Ochrobactrum anthropi NCIMB41200, or Clostridium kluyveri DSM555.
- 5. Process according to claim 1 or 2, characterized in that the enzyme having α,β-enoate reductase activity has aerostable α,β-enoate reductase activity and is an enzyme originating from a microorganism from the group of species of Agrobacterium sp., Burkholderia sp., Escherichia sp., Pseudomonas sp., Salmonella sp., Shigella sp., Yersinia sp., and Vibrio sp.
- 6. Process according to claim 5, characterized in that the enzyme having aerostable α,β-enoate reductase activity is an enzyme originating from an Escherichia coli species.
- 7. Process according to claim 6, characterized in that the enzyme having aerostable α,β-enoate reductase activity is an enzyme originating from from Escherichia coli K12.

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- Process according to any of claims 1-7, characterized in that
 6-aminohex-2-enoic acid is being converted into 6-amino caproic acid at a pH in the range of from 3 to 9.
- 9. Process according to claim 8, characterized in that, the pH is in the range of from 4 to 8.
- 10. Process according to claim 9, characterized in that the pH is in the range of from 5 to 8.
- 11. Process according to claim 8, characterized in that, the pH is in the range of from 5.5 to 7 under anaerobic conditions and of from 6.5 to 8 under aerobic conditions.
- 12. Process according to any of claims 1-11, characterized in that the process is carried out in a host organism selected from the group of genera consisting of Aspergillus, Bacillus, Corynebacterium, Escherichia and Pichia.
- 13. Process according to claim 12, characterized in that the process is carried out in a host organism selected from the group of *Escherichia coli*, *Bacillus*, *Corynebacterium glutarnicum*, *Aspergillus niger* or *Pichia pastoris* host organisms.
- 14. Process according to claim 12 or 13, characterized in that in the host organism an α,β-enoate reductase gene encoding an enzyme having α,β-enoate reductase activity towards molecules containing an α,β-enoate group and a primary amino group is cloned and expressed.
- 15. An Escherichia coli host cell wherein the α,β-enoate reductase gene from Ochrobactrum anthropi NCIMB41200, or from Acremonium strictum CBS114157 is cloned and expressed.
- 16. A Bacillus host cell wherein the α,β-enoate reductase gene from Moorella thermoacetica DSM1974, or from Clostridium tyrobutyricum DSM1460, or from Ochrobactrum anthropi NCIMB41200, or from Acremonium strictum CBS114157 is cloned and expressed.
- 17. A Corynebacterium glutamicum host ceil wherein the α,β-enoate reductase gene from Moorella thermoacetica DSM1974, or from Clostridium tyrobutyricum DSM1460, or from Ochrobactrum anthropi NCIMB41200, or from Acremonium strictum CBS114157 is cloned and expressed.
- An Aspergillus niger host cell wherein the α,β-enoate reductase gene from Acremonium strictum CBS114157, or from Moorella thermoacetica DSM1974, or from Clostridium tyrobutyricum DSM1460, or from Ochrobactrum anthropi NCIMB41200 is cloned and expressed.

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- 19. A Pichia pastoris host cell wherein the α,β-enoate reductase gene from Acremonium strictum CBS114157, or from Moorella thermoacetica DSM1974, or from Clostridium tyrobutyricum DSM1460, or from Ochrobactrum anthropi NCIMB41200 is cloned and expressed.
- A host cell selected from the group of Aspergillus, Bacillus,

 Corynebacterium, and Pichia host cells, in which the aerostable α,β-enoate reductase gene nemA from E. coli K12 is cloned and expressed.
- 21. Process for precursor fermentation of 6-amino caproic acid starting either from 6-aminohex-2-enoic acid (6-AHEA) or from 6-amino-2-hydroxyhexanoic acid (6-AHHA), and applying at least an enzymatic step with an enzyme having α,β-enoate reductase activity towards molecules containing an α,β-enoate group and a primary amino group, in particular with an enzyme having α,β-enoate reductase activity towards 6-aminohex-2-enoic acid.
- 22. Process according to claim 21, characterized in that the process is performed in a microorganism wherein 6-aminohex-2-enoic acid is being formed *in vivo*.
- 23. Process according to claim 22, characterized in that 6-aminohex-2-enoic acid is being formed *in vivo* from solutions or slurries containing a suitable carbon source.
- 24. Biochemically produced 6-aminohex-2-enoic acid, having a ¹²C versus ¹³C versus ¹⁴C isotope ratio of about the same value as occurring in environmental carbon dioxide.
- 25. Biochemically produced 6-amino-hexanoic acid having a ¹²C versus ¹³C versus ¹⁴C isotope ratio of about the same value as occurring in environmental carbon dioxide.
- 26. ε-Caprolactam produced from biochemically produced 6-aminohex-2-enoic acid or 6-amino-hexanoic acid, and having a ¹²C versus ¹³C versus ¹⁴C isotope ratio of about the same value as occurring in environmental carbon dioxide.
- 27. Nylon-6 and other derivatives produced from any of the biochemically produced products of claims 24 or 25, or from ε-caprolactam according to claim 26, and having a ¹²C versus ¹³C versus ¹⁴C isotope ratio of about the same value as occurring in environmental carbon dioxide.